



COMMENTARY

Will you still need me (Ca²⁺, TnT, and DHPR), will you still cleave me (calpain), when I'm 64?

José Renato Pinto,¹ Judy Muller-Delp¹ and P. Bryant Chase²

¹Department of Biomedical Sciences, The Florida State University College of Medicine, 1115 West Call Street, Tallahassee, FL 32306-4300, USA

²Department of Biological Science, The Florida State University, 81 Chieftain Way, Tallahassee, FL 32306-4370, USA

Of the many cellular and molecular hallmarks that are broadly associated with physiological decline during aging (López-Otín *et al.*, 2013), loss of muscular strength in vertebrates is particularly problematic because in humans it is a better predictor of morbidity and mortality than loss of muscle mass (Newman *et al.*, 2006). Human cohort studies indicate that with both aging and disease, muscular strength is lost more rapidly than muscle mass (Goodpaster *et al.*, 2006). The mechanistic changes that underlie age-related loss of muscular strength, however, have been more elusive to identify than the mechanisms of age-related sarcopenia. Age-induced loss of muscular strength has been a topic of sustained debate. Despite a number of plausible hypotheses and clever experimental designs, these earlier studies were unable to dissect the primary mechanism(s) responsible for the reduction in specific force (when the force is normalized to the cross-sectional area) with aging (Phillips *et al.*, 1993; Brooks & Faulkner, 1994; Faulkner *et al.*, 2007). More recently, the Delbono group demonstrated that decreased expression of the voltage sensor Ca²⁺ channel α 1 subunit (Cav1.1)—also known as the dihydropyridine receptor (DHPR) in the skeletal muscle excitation–contraction coupling literature—is associated with the loss of skeletal muscle strength with aging (Delbono *et al.*, 2007; Taylor *et al.*, 2009). They also showed that Cav1.1 expression levels can be regulated by different mechanisms, which are not related to gene transcription or mRNA expression (Delbono *et al.*, 2007; Taylor *et al.*, 2009).

In the paper 'Calpain inhibition rescues troponin T3 fragmentation, increases Cav1.1, and enhances skeletal muscle force in aging sedentary mice', Zhang *et al.* report a novel finding that TnT3 regulates Cav1.1 expression in skeletal muscle fibers and that calpain-mediated fragmentation of TnT3 is associated with Cav1.1 downregulation in old mice (Zhang *et al.*, 2016). Delbono and colleagues have described how the age-induced decrease in Cav1.1 levels leads to uncoupling of the type 1 ryanodine receptor (RyR1), which potentially decreases the amount of activating Ca²⁺ released by the sarcoplasmic reticulum during contraction (Fig. 1) (Delbono, 2011; Hernandez-Ochoa *et al.*, 2015; Lee *et al.*, 2015). The coupling between Cav1.1 in the sarcolemma and RyR1 in the sarcoplasmic reticulum membrane has also been recently shown to be modulated by protein Stac3, a novel mechanism for the modulation of excitation–contraction that does not involve changes in the cellular level of Cav1.1 (Polster *et al.*, 2016). Importantly, Zhang *et al.* also report that calpain-mediated TnT3 fragmentation and related downregulation of

Cav1.1 expression can be prevented by administration of a calpain inhibitor, BDA-410, to old mice. These results suggest that calpain activity and reduction in TnT3 fragmentation are potential therapeutic targets for prevention and/or amelioration of age-related loss of muscular strength.

TnT3 is one of the three polypeptides that comprise the troponin complex in skeletal muscle. The function of the sarcomeric troponin complex in the context of striated muscle regulation has been extensively studied (Farah & Reinach, 1995; Gordon *et al.*, 2000; Vinogradova *et al.*, 2005). In addition to cytoplasmic localization in thin filaments of the sarcomere, TnT and troponin I (TnI)—along with troponin C (TnC), tropomyosin, and actin—are present in the nuclei of striated muscle cells (Asumda & Chase, 2012; Chase *et al.*, 2013; Zhang *et al.*, 2013a,b). However, the role of troponin subunits in the striated muscle nucleus is still under investigation and little is known. A mutant human cardiac TnT (cTnT-R173W) associated with dilated cardiomyopathy (DCM) accumulates in the nuclei of iPSC-derived cardiomyocytes and upregulates phosphodiesterase (PDE) 2A and 3A activities via modulation of epigenetic factors (Wu *et al.*, 2015). Increased PDE activity is correlated with increased cAMP levels and impaired β -adrenergic signaling, which are hallmarks of the disease in patients with DCM and which compound the detrimental effects of the TnT mutation on sarcomere contractility. In the paper by Zhang *et al.* (2016), TnT3—the TnT isoform found in fast-twitch skeletal muscle fibers—is reported to bind to the promoter region of the *Cacna1s* (gene encoding Cav1.1) and regulate its transcription levels in skeletal muscle fibers.

In Zhang *et al.*, the authors demonstrated that the expression of Cav1.1 is coupled to the expression of TnT3 and that TnT3—but not TnI, TnC, or tropomyosin—binds specifically to the promoter region of *Cacna1s*. Then they used SDS-PAGE to separate nuclear protein extracts obtained from old mice (23–25 months) and identified a fragment of TnT3. Mass spectrometry analysis indicated that cleavage sites of the TnT3 fragments corresponded to sequences targeted by calpain. Subsequently, the authors elegantly showed that old mice treated with BDA-410 (a synthetic Leu–Leu peptidomimetic that inhibits cysteine proteases) daily for 21 days displayed increased absolute and specific forces at all frequencies of stimulation tested in intact soleus muscle preparations, without changes in CSA. The endurance capacity tested *in vivo* and *ex vivo* was not improved by the BDA-410 treatment. Furthermore, they showed that BDA-410 does not affect the composition of soleus and EDL muscles; for example, the levels of titin and myosin heavy chain (MHC) were not altered as well as MHC isoform distribution in these two muscles. Interestingly, BDA-410 treatment stabilized the nuclear full-length TnT3 and decreased the levels of fragmented TnT3, which consequently increased the amount of Cav1.1 in skeletal muscle cells (Fig. 1) (Zhang *et al.*, 2016).

Current therapeutic approaches that are being tested to improve skeletal muscle function in the context of aging and disease include myostatin inhibition, hormone therapy, and troponin activation, among others (Jasuja & Lebrasseur, 2014). Calpain activity has been linked to several disease conditions in striated muscle (Jia *et al.*, 2001;

Correspondence

José Renato Pinto, Department of Biomedical Sciences, Florida State University College of Medicine, 1115 West Call Street, Tallahassee, FL 32306-4300, USA. Tel.: +1 850 645 0016; fax: +1 850 644 5781; e-mail: jose.pinto@med.fsu.edu

Accepted for publication 19 November 2016

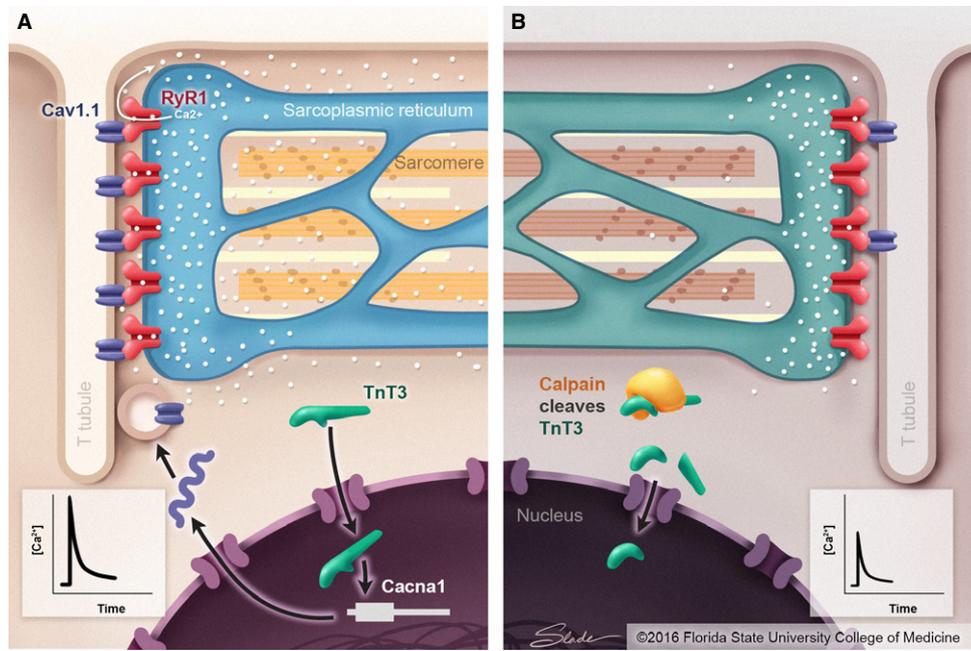


Fig. 1 Illustration of the molecular mechanism by which TnT3 regulates expression of Cav1.1 in skeletal muscle. (A) Young skeletal muscle cell and (B) aged skeletal muscle cell.

Witt *et al.*, 2004; Zhang *et al.*, 2006; Patterson *et al.*, 2011), and inhibition of its activity has been suggested as a therapeutic strategy (Carragher, 2006), leading to the identification of BDA-410—the calpain inhibitor used by Zhang *et al.*—and the search for other small molecule inhibitors (Xu *et al.*, 2013). The report by Zhang *et al.* demonstrates a role for nuclear TnT3 regulating *Cacna1s* transcription and Cav1.1 expression, and it also indicates that inhibition of skeletal muscle calpain activity may be an effective new therapeutic strategy to diminish the deleterious effects of aging on muscle function. Another potential topic of investigation is whether resistance training, which is known to effectively increase strength through mechanisms other than hypertrophy in aged muscle (Frontera *et al.*, 1988), exerts its beneficial effect on muscle performance by decreasing nuclear fragmented TnT3. This study by Zhang *et al.* provides both important mechanistic data for understanding the regulation of Cav1.1 in aged skeletal muscle and direction for future interventional studies in the aging population.

Funding

National Heart, Lung and Blood Institute of the National Institutes of Health Grant HL128683 to JRP.

Conflict of interest

None declared.

References

- Asumda FZ, Chase PB (2012) Nuclear cardiac troponin and tropomyosin are expressed early in cardiac differentiation of rat mesenchymal stem cells. *Differentiation* **83**, 106–115.
- Brooks SV, Faulkner JA (1994) Skeletal muscle weakness in old age: underlying mechanisms. *Med. Sci. Sports Exerc.* **26**, 432–439.
- Carragher NO (2006) Calpain inhibition: a therapeutic strategy targeting multiple disease states. *Curr. Pharm. Des.* **12**, 615–638.
- Chase PB, Szczypinski MP, Soto EP (2013) Nuclear tropomyosin and troponin in striated muscle: new roles in a new locale? *J. Muscle Res. Cell Motil.* **34**, 275–284.
- Delbono O (2011) Expression and regulation of excitation-contraction coupling proteins in aging skeletal muscle. *Curr. Aging Sci.* **4**, 248–259.
- Delbono O, Xia J, Treves S, Wang ZM, Jimenez-Moreno R, Payne AM, Messi ML, Briguet A, Schaerer F, Nishi M, Takeshima H, Zorzato F (2007) Loss of skeletal muscle strength by ablation of the sarcoplasmic reticulum protein JP45. *Proc. Natl. Acad. Sci. U.S.A.* **104**, 20108–20113.
- Farah CS, Reinach FC (1995) The troponin complex and regulation of muscle contraction. *FASEB J.* **9**, 755–767.
- Faulkner JA, Larkin LM, Clafflin DR, Brooks SV (2007) Age-related changes in the structure and function of skeletal muscles. *Clin. Exp. Pharmacol. Physiol.* **34**, 1091–1096.
- Frontera WR, Meredith CN, Reilly KP, Knuttgen HG, Evans WJ (1988) Strength conditioning in older men: skeletal muscle hypertrophy and improved function. *J. Appl. Physiol.* **64**, 1038.
- Goodpaster BH, Park SW, Harris TB, Kritchevsky SB, Nevitt M, Schwartz AV, Simonsick EM, Tylavsky FA, Visser M, Newman AB (2006) The loss of skeletal muscle strength, mass, and quality in older adults: the health, aging and body composition study. *J. Gerontol. A Biol. Sci. Med. Sci.* **61**, 1059–1064.
- Gordon AM, Homsher E, Regnier M (2000) Regulation of contraction in striated muscle. *Physiol. Rev.* **80**, 853–924.
- Hernandez-Ochoa EO, Pratt SJ, Lovering RM, Schneider MF (2015) Critical role of intracellular RyR1 calcium release channels in skeletal muscle function and disease. *Front. Physiol.* **6**, 420.
- Jasuja R, Lebrasseur NK (2014) Regenerating skeletal muscle in the face of aging and disease. *Am. J. Phys. Med. Rehabil.* **93**, S88–S96.
- Jia Z, Petrounevitch V, Wong A, Moldoveanu T, Davies PL, Elce JS, Beckmann JS (2001) Mutations in calpain 3 associated with limb girdle muscular dystrophy: analysis by molecular modeling and by mutation in m-calpain. *Biophys. J.* **80**, 2590–2596.
- Lee CS, Dagnino-Acosta A, Yarotsky V, Hanna A, Lyfenko A, Knoblauch M, Georgiou DK, Poche RA, Swank MW, Long C, Ismailov II, Lanner J, Tran T, Dong K, Rodney GG, Dickinson ME, Beeton C, Zhang P, Dirksen RT, Hamilton SL (2015) Ca(2+) permeation and/or binding to Cav1.1 fine-tunes skeletal muscle Ca(2+) signaling to sustain muscle function. *Skelet Muscle* **5**, 4.
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G (2013) The hallmarks of aging. *Cell* **153**, 1194–1217.

- Newman AB, Kupelian V, Visser M, Simonsick EM, Goodpaster BH, Kritchevsky SB, Tylavsky FA, Rubin SM, Harris TB (2006) Strength, but not muscle mass, is associated with mortality in the health, aging and body composition study cohort. *J. Gerontol. A Biol. Sci. Med. Sci.* **61**, 72–77.
- Patterson C, Portbury AL, Schisler JC, Willis MS (2011) Tear me down: role of calpain in the development of cardiac ventricular hypertrophy. *Circ. Res.* **109**, 453–462.
- Phillips SK, Wiseman RW, Woledge RC, Kushmerick MJ (1993) Neither changes in phosphorus metabolite levels nor myosin isoforms can explain the weakness in aged mouse muscle. *J. Physiol.* **463**, 157–167.
- Polster A, Nelson BR, Olson EN, Beam KG (2016) Stac3 has a direct role in skeletal muscle-type excitation-contraction coupling that is disrupted by a myopathy-causing mutation. *Proc. Natl. Acad. Sci. U.S.A.* **113**, 10986–10991.
- Taylor JR, Zheng Z, Wang ZM, Payne AM, Messi ML, Delbono O (2009) Increased CaVbeta1A expression with aging contributes to skeletal muscle weakness. *Aging Cell* **8**, 584–594.
- Vinogradova MV, Stone DB, Malanina GG, Karatzaferi C, Cooke R, Mendelson RA, Fletterick RJ (2005) Ca(2+)-regulated structural changes in troponin. *Proc. Natl. Acad. Sci. U.S.A.* **102**, 5038–5043.
- Witt CC, Ono Y, Puschmann E, McNabb M, Wu Y, Gotthardt M, Witt SH, Haak M, Labeit D, Gregorio CC, Sorimachi H, Granzier H, Labeit S (2004) Induction and myofibrillar targeting of CARP, and suppression of the Nkx2.5 pathway in the MDM mouse with impaired titin-based signaling. *J. Mol. Biol.* **336**, 145–154.
- Wu H, Lee J, Vincent LG, Wang Q, Gu M, Lan F, Churko JM, Sallam KI, Matsa E, Sharma A, Gold JD, Engler AJ, Xiang YK, Bers DM, Wu JC (2015) Epigenetic regulation of phosphodiesterases 2A and 3A underlies compromised β -adrenergic signaling in an iPSC model of dilated cardiomyopathy. *Cell Stem Cell* **17**, 89–100.
- Xu C, Tabebordbar M, Iovino S, Ciarlo C, Liu J, Castiglioni A, Price E, Liu M, Barton ER, Kahn CR, Wagers AJ, Zon LI (2013) A zebrafish embryo culture system defines factors that promote vertebrate myogenesis across species. *Cell* **155**, 909–921.
- Zhang Z, Biesiadecki BJ, Jin J-P (2006) Selective deletion of the NH₂-terminal variable region of cardiac troponin T in ischemia reperfusion by myofibril-associated μ -calpain cleavage. *Biochemistry* **45**, 11681–11694.
- Zhang T, Birbrair A, Delbono O (2013a) Nonmyofibril-associated troponin T3 nuclear and nucleolar localization sequence and leucine zipper domain mediate muscle cell apoptosis. *Cytoskeleton* **70**, 134–147.
- Zhang T, Birbrair A, Wang ZM, Taylor J, Messi ML, Delbono O (2013b) Troponin T nuclear localization and its role in aging skeletal muscle. *Age (Dordr)* **35**, 353–370.
- Zhang T, Pereyra AS, Wang ZM, Birbrair A, Reisz JA, Files DC, Purcell L, Feng X, Messi ML, Feng H, Chalovich J, Jin JP, Furdulj C, Delbono O (2016) Calpain inhibition rescues troponin T3 fragmentation, increases Cav1.1, and enhances skeletal muscle force in aging sedentary mice. *Aging Cell* **15**, 488–498.